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(54) Title: MATRIX-REMODELING GENES

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(57) Abstract

The invention provides novel matrix-remodeling genes and polypeptides encoded by those genes. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating or preventing diseases associated with matrix remodeling.

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MATRIX-REMODELING GENES

TECHNICAL FIELD

The invention relates to novel matrix-remodeling genes identified by their coexpression with known matrix-remodeling genes. The invention also relates to the use of these biomolecules in diagnosis, prognosis, prevention, treatment, and evaluation of therapies for diseases, particularly diseases associated with matrix-remodeling such as cancer, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis, atherosclerosis, fibrosis, and ulceration.

BACKGROUND OF THE INVENTION

Matrix remodeling is associated with the construction, destruction, and reorganization of extracellular matrix components and is essential in normal cellular functions and also in many disease processes. These disease processes include metastatic cancer, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis, atherosclerosis, fibrosis, and ulceration (Alexander and Werb (1991) In: Cell Biology of Extracellular Matrix, Plenum Press, New York NY, pp. 255-302; Schuppan et al. (1993) In: Extracellular Matrix, Marcel Dekker, New York NY, pp. 201-254; Zvibel and Kraft (1993) In: Extracellular Matrix, Marcel Dekker, New York NY, pp. 559-580; Shanahan et al. (1994) J Clin Invest 93:2393-402; Kielty and Shuttleworth (1995) Int J Biochem Cell Biol 27:747-60; Bitar and Labbad (1996) J Surg Res 61:113-9; Dourado et al. (1996) Osteoarthritis Cartilage 4:187-96; Grant et al. (1996) Regul. Pept. 67:137-44; Gunja-Smith et al. (1996) Am J Pathol 148:1639-48; Alcolado et al. (1997) Clin. Sci 92:103-12; Cs-Szabo et al. (1997) Arthritis Rheum 40:1037-45; Hayward and Brock (1997) Hum Mutat 10:415-23; Ledda et al. (1997) J Invest Dermatol 108:210-4; Hayashido et al. (1998) Int J Cancer 75:654-8; Ito et al. (1998) Kidney Int 53:853-61; Nelson et al. (1998) Cancer Res 58:232-6).

Many genes that participate in and regulate matrix remodeling are known, but many remain to be identified. Identification of currently unknown genes will provide new diagnostic and therapeutic targets. In addition, these genes will provide new opportunities for therapeutic tissue engineering--the use of drugs or biologicals to direct the creation of new tissues such as skin, pancreas, or liver that can replace tissues lost to disease or trauma.

The present invention provides new compositions that are useful for diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases associated with matrix remodeling. We have implemented a method for analyzing gene expression patterns and have identified 20 novel matrix-remodeling genes by their coexpression with known matrix-remodeling genes.

SUMMARY OF THE INVENTION

In one aspect, the invention provides for a substantially purified polynucleotide comprising a gene that is coexpressed with one or more known matrix-remodeling genes in a plurality of biological samples. Preferably, each known matrix-remodeling gene is selected from the group consisting of osteonectin (BM-40), chondroitin/dermatan sulfate proteoglycans (C/DSPG), collagen I, II, II, and IV, connective tissue growth factor (CTGF), fibrillin, fibronectins, fibronectin receptor (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases (MMP), and tissue inhibitors of matrix metalloproteinase 1, 2, and 3 (TIMP 1, 2, and 3). Preferred embodiments are (a) a polynucleotide sequence selected from the group consisting of SEQ ID NOs:1-20; (b) a polynucleotide sequence which encodes a polypeptide sequence of SEQ ID NOs:21, 22, and 23; (c) a polynucleotide sequence having at least 70% identity to the polynucleotide sequence of (a) or (b); (d) a polynucleotide sequence comprising at least 18 sequential nucleotides of the polynucleotide sequence of (a), (b), or (c); (e) a polynucleotide which hybridizes under stringent conditions to the polynucleotide of (a), (b), (c), or (d); or (f) a polynucleotide sequence which is complementary to the polynucleotide sequence of (a), (b), (c), (d) or (e). Furthermore, the invention provides an expression vector comprising any of the above described polynucleotides and host cells comprising the expression vector. Still further, the invention provides a method for treating or preventing a disease or condition associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a sample comprising administering to a subject in need the above-described polynucleotides in an amount effective for treating or preventing said disease.

In a second aspect, the invention provides a substantially purified polypeptide comprising the gene product of a gene that is coexpressed with one or more known matrix-

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remodeling genes in a plurality of biological samples. The known matrix-remodeling gene may be selected from the group consisting of osteonectin (BM-40), chondroitin/dermatan sulfate proteoglycans (C/DSPG), collagen I, II, II, and IV, connective tissue growth factor (CTGF), fibrillin, fibronectins, fribonectin receptors (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases (MMP), and tissue inhibitors of matrix metalloproteinase 1. 2, and 3 (TIMP 1, 2, and 3). Preferred embodiments are polypeptides comprising (a) the polypeptide sequence of SEQ ID NO:21, 22, or 23; (b) a polypeptide sequence having at least 85% identity to the polypeptide sequence of (a); and (c) a polypeptide sequence comprising at least 6 sequential amino acids of the polypeptide sequence of (a) or (b). Additionally, the invention provides antibodies that bind specifically to any of the above described polypeptides and a method for treating or preventing a disease or condition associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a sample comprising administering to a subject in need such an antibody in an amount effective for treating or preventing said disease.

In another aspect, the invention provides a pharmaceutical composition comprising the polynucleotide of claim 2 or the polypeptide of claim 3 in conjunction with a suitable pharmaceutical carrier or a method for treating or preventing a disease or condition associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a sample comprising administering to a subject in need such compositioning in an amount effective for treating or preventing said disease.

In yet a further aspect, the invention provides a method for diagnosing a disease or condition associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a sample, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin (BM-40), chondroitin/dermatan sulfate proteioglycans (C/DSPG), collagen I, II, II, and IV, connective tissue growth factor (CTGF), fibrillin, fibronectins, fibronectin receptor (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases (MMP), and tissue inhibitors of matrix metalloproteinase 1, 2, and 3 (TIMP)

1, 2, and 3). The method comprises the steps of (a) providing the sample comprising one of more of said coexpressed genes; (b) hybridizing the polynucleotide of the coexpressed genes under conditions effective to form one or more hybridization complexes; and (c) detecting the hybridization complexes, wherein the altered level of one or more of the hybridization
 5 complexes in a diseased sample compared with the level of hybridization complexes in a non-diseased sample correlates with the presence of the disease or condition in the sample.

BRIEF DESCRIPTION OF THE SEQUENCE LISTING

The Sequence Listing provides exemplary matrix-remodeling-associated sequences including polynucleotide sequences, SEQ ID NOs:1-20, and polypeptide sequences, SEQ ID NOs:21-23. Each sequence is identified by a sequence identification number (SEQ ID NO) and by the Incyte Clone number from which the sequence was first identified.

DESCRIPTION OF THE INVENTION

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include the plural reference unless the context clearly dictates otherwise.

Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a

reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

DEFINITIONS

"NSEQ" refers generally to a polynucleotide sequence of the present invention, including SEQ ID NOs:1-20. "PSEQ" refers generally to a polypeptide sequence of the present invention, including SEQ ID NOs:21-23.

A "variant" refers to either a polynucleotide or a polypeptide whose sequence diverges from SEQ ID NOs:1-20 or SEQ ID NOs:21-23, respectively. Polynucleotide sequence divergence may result from mutational changes such as deletions, additions, and substitutions of one or more nucleotides; it may also occur because of differences in codon usage. Each of these types of changes may occur alone, or in combination, one or more times in a given sequence. Polypeptide variants include sequences that possess at least one structural or functional characteristic of SEQ ID NOs:21-23.

A "fragment" can refer to a nucleic acid sequence that is preferably at least 20 nucleic acids in length, more preferably 40 nucleic acids, and most preferably 60 nucleic acids in length, and encompasses, for example, fragments consisting of nucleic acids 1-50 or 200-500

of SEQ ID NOs:1-20. A "fragment" can also refer to polypeptide sequences which are preferably at least 5 to about 15 amino acids in length, most preferably at least 10 amino acids long, and which retain some biological activity or immunological activity of, for example, a sequence selected from SEQ ID NOs:21-23.

"Gene" or "gene sequence" refers to the partial or complete coding sequence of a transcript. The term also refers to sequences corresponding to 5' or 3' untranslated regions or 5' or 3' untranslated regions including partial or complete coding sequences of a gene.

Typically, the novel gene sequences may or may not be homolgous to annotated sequences found in public or private databases. The gene may be in a sense or antisense (complementary) orientation.

"Known matrix-remodeling gene" refers to a gene sequence which has been previously identified as useful in the diagnosis, treatment, prognosis, or prevention of diseases associated with matrix remodeling. Typically, this means that the known matrix-remodeling gene is expressed at higher levels in tissue abundant in known matrix-remodeling transcripts when compared with other tissue.

"Matrix-remodeling gene" refers to a gene sequence whose expression pattern is similar to that of the known matrix-remodeling genes and which are useful in the diagnosis, treatment, prognosis, or prevention of diseases associated with matrix remodeling. The gene sequences can also be used in the evaluation of therapies for cancer.

"Substantially purified" refers to a nucleic acid or an amino acid sequence that is removed from its natural environment and is isolated or separated, and is at least about 60% free, preferably about 75% free, and most preferably about 90% free from other components with which it is naturally present.

THE INVENTION

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The present invention encompasses a method for identifying biomolecules that are associated with a specific disease, regulatory pathway, subcellular compartment, cell type, tissue type, or species. In particular, the method identifies gene sequences useful in diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases associated with matrix-remodeling, particularly, cancer, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis, atherosclerosis, fibrosis, and ulceration.

The method provides first identifying polynucleotides that are expressed in a plurality

of cDNA libraries. The identified polynucleotides include genes of known function, genes known to be specifically expressed in a specific disease process, subcellular compartment, cell type, tissue type, or species. Additionally, the polynucleotides include genes of unknown function. The expression patterns of the known genes are then compared with those of the genes of unknown function to determine whether a specified coexpression probability threshold is met. Through this comparison, a subset of the polynucleotides for unknown function genes having a high coexpression probability with the known genes can be identified. The high coexpression probability correlates with a particular coexpression probability threshold which is less than 0.001, and more preferably less than 0.00001.

The polynucleotides originate from cDNA libraries derived from a variety of sources including, but not limited to, eukaryotes such as human, mouse, rat, dog, monkey, plant, and yeast and prokaryotes such as bacteria and viruses. These polynucleotides can also be selected from a variety of sequence types including, but not limited to, expressed sequence tags (ESTs), assembled polynucleotide sequences, full length gene coding regions, introns, 15 regulatory sequences, 5' untranslated regions, and 3' untranslated regions. To have statistically significant analytical results, the polynucleotides need to be expressed in at least three cDNA libraries.

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The cDNA libraries used in the coexpression analysis of the present invention can be obtained from blood vessels, heart, blood cells, cultured cells, connective tissue, epithelium, islets of Langerhans, neurons, phagocytes, biliary tract, esophagus, gastrointestinal system, liver, pancreas, fetus, placenta, chromaffin system, endocrine glands, ovary, uterus, penis, prostate, seminal vesicles, testis, bone marrow, immune system, cartilage, muscles, skeleton, central nervous system, ganglia, neuroglia, neurosecretory system, peripheral nervous system, bronchus, larynx, lung, nose, pleurus, ear, eye, mouth, pharynx, exocrine glands, bladder, 25 kidney, ureter, and the like. The number of cDNA libraries selected can range from as few as 20 to greater than 10,000. Preferably, the number of the cDNA libraries is greater than 500.

In a preferred embodiment, gene sequences are assembled to reflect related sequences, such as assembled sequence fragments derived from a single transcript. Assembly of the polynucleotide sequences can be performed using sequences of various types including, but 30 not limited to, ESTs, extensions, or shotgun sequences. In a most preferred embodiment, the polynucleotide sequences are derived from human sequences that have been assembled using

the algorithm disclosed in "Database and System for Storing, Comparing and Displaying Related Biomolecular Sequence Information", Lincoln et al., Serial No:60/079,469, filed March 26, 1998, incorporated herein by reference.

Experimentally, differential expression of the polynucleotides can be evaluated by

5 methods including, but not limited to, differential display by spatial immobilization or by gel
electrophoresis, genome mismatch scanning, representational difference analysis, and
transcript imaging. Additionally, differential expression can be assessed by microarray
technology. These methods may be used alone or in combination.

Known matrix-remodeling genes can be selected based on the use of the genes as

diagnostic or prognostic markers or as therapeutic targets for diseases associated with matrix
remodeling, such as cancer, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis,
atherosclerosis, fibrosis, and ulceration. Preferably, the known matrix-remodeling genes
include osteonectin (BM-40), chondroitin/dermatan sulfate proteioglycans (C/DSPG),
collagen I, II, II, and IV, connective tissue growth factor (CTGF), fibrillin, fibronectins,
fibronectin receptor (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular
matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding
protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases
(MMP), tissue inhibitors of matrix metalloproteinase 1, 2, and 3 (TIMP 1, 2, and 3), and the
like.

The procedure for identifying novel genes that exhibit a statistically significant coexpression pattern with known matrix-remodeling genes is as follows. First, the presence or absence of a gene sequence in a cDNA library is defined: a gene is present in a cDNA library when at least one cDNA fragment corresponding to that gene is detected in a cDNA sample taken from the library, and a gene is absent from a library when no corresponding cDNA fragment is detected in the sample.

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Second, the significance of gene coexpression is evaluated using a probability method to measure a due-to-chance probability of the coexpression. The probability method can be the Fisher exact test, the chi-squared test, or the kappa test. These tests and examples of their applications are well known in the art and can be found in standard statistics texts (Agresti, A (1990) Categorical Data Analysis, John Wiley & Sons, New York NY; Rice, JA (1988) Mathematical Statistics and Data Analysis, Duxbury Press, Pacific Grove CA). A Bonferroni

correction (Rice, <u>supra</u>, page 384) can also be applied in combination with one of the probability methods for correcting statistical results of one gene versus multiple other genes. In a preferred embodiment, the due-to-chance probability is measured by a Fisher exact test, and the threshold of the due-to-chance probability is set to less than 0.001, more preferably less than 0.0001.

To determine whether two genes, A and B, have similar coexpression patterns, occurrence data vectors can be generated as illustrated in Table 1, wherein a gene's presence is indicated by a one and its absence by a zero. A zero indicates that the gene did not occur in the library, and a one indicates that it occurred at least once.

Table 1. Occurrence data for genes A and B

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	Library 1	Library 2	Library 3	 Library N
gene A	1	1	0	 0
gene B	1	0	1	 0

15 For a given pair of genes, the occurrence data in Table 1 can be summarized in a 2 x 2 contingency table.

Table 2. Contingency table for co-occurrences of genes A and B

	Gene A present	Gene A absent	Total
Gene B present	8	2	10
Gene B absent	2	18	20
Total	10	20	30

Table 2 presents co-occurrence data for gene A and gene B in a total of 30 libraries. Both gene A and gene B occur 10 times in the libraries. Table 2 summarizes and presents 1) the number of times gene A and B are both present in a library, 2) the number of times gene A and B are both absent in a library, 3) the number of times gene A is present while gene B is absent, and 4) the number of times gene B is present while gene A is absent. The upper left entry is the number of times the two genes co-occur in a library, and the middle right entry is the number of times neither gene occurs in a library. The off diagonal entries are the number of times one gene occurs while the other does not. Both A and B are present eight times and absent 18 times, gene A is present while gene B is absent two times, and gene B is present while gene A

is absent two times. The probability ("p-value") that the above association occurs due to chance as calculated using a Fisher exact test is 0.0003. Associations are generally considered significant if a p-value is less than 0.01 (Agresti, supra; Rice, supra).

This method of estimating the probability for coexpression of two genes makes several assumptions. The method assumes that the libraries are independent and are identically sampled. However, in practical situations, the selected cDNA libraries are not entirely independent because more than one library may be obtained from a single patient or tissue, and they are not entirely identically sampled because different numbers of cDNAs may be sequenced from each library (typically ranging from 5,000 to 10,000 cDNAs per library). In addition, because a Fisher exact coexpression probability is calculated for each gene versus 41,419 other genes, a Bonferroni correction for multiple statistical tests is necessary.

Using the method of the present invention, we have identified 20 novel genes that exhibit strong association, or coexpression, with known genes that are matrix-remodeling-specific. These known matrix-remodeling genes include osteonectin (BM-40),

15 chondroitin/dermatan sulfate proteioglycans (C/DSPG), collagen I, II, II, and IV, connective tissue growth factor (CTGF), fibrillin, fibronectins, fibronectin receptor (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases (MMP), and tissue inhibitors of matrix metalloproteinase 1, 2, and 3 (TIMP 1, 2, and 3). The results presented in Tables 5 and 6 show that the expression of the 20 novel genes have direct or indirect association with the expression of known matrix-remodeling genes. Therefore, the novel genes can potentially be used in diagnosis, treatment, prognosis, or prevention of diseases associated with matrix remodeling.

25 Further, the gene products of the 20 novel genes are potential therapeutic proteins and targets of therapeutics against diseases associated with matrix remodeling.

Therefore, in one embodiment, the present invention encompasses a polynucleotide sequence comprising the sequence of SEQ ID NOs:1-20. These 20 polynucleotides are shown by the method of the present invention to have strong coexpression association with known matrix-remodeling genes and with each other. The invention also encompasses a variant of the polynucleotide sequence, its complement, or 18 consecutive nucleotides of a sequence

provided in the above described sequences. Variant polynucleotide sequences typically have at least about 70%, more preferably at least about 85%, and most preferably at least about 95% polynucleotide sequence identity to NSEQ.

One preferred method for identifying variants entails using NSEQ and/or PSEQ

sequences to search against the GenBank primate (pri), rodent (rod), and mammalian (mam), vertebrate (vrtp), and eukaryote (eukp) databases, SwissProt, BLOCKS (Bairoch et al. (1997) Nucleic Acids Res 25:217-221), PFAM, and other databases that contain previously identified and annotated motifs, sequences, and gene functions. Methods that search for primary sequence patterns with secondary structure gap penalties (Smith et al. (1992) Protein

Engineering 5:35-51) as well as algorithms such as BLAST (Basic Local Alignment Search Tool; Altschul (1993) J Mol Evol 36:290-300; and Altschul et al. (1990) J Mol Biol 215:403-410), BLOCKS (Henikoff and Henikoff (1991) Nucleic Acids Res 19:6565-6572), Hidden Markov Models (HMM; Eddy (1996) Cur Opin Str Biol 6:361-365; Sonnhammer et al. (1997) Proteins 28:405-420), and the like, can be used to manipulate and analyze nucleotide and amino acid sequences. These databases, algorithms and other methods are well known in the art and are described in Ausubel et al. (1997; Short Protocols in Molecular Biology, John Wiley & Sons, New York NY) and in Meyers (1995; Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853).

Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to SEQ ID NOs:1-20, and fragments thereof under stringent conditions. Stringent conditions can be defined by salt concentration, temperature, and other chemicals and conditions well known in the art. In particular, stringency can be increased by reducing the concentration of salt, or raising the hybridization temperature. Varying additional parameters, such as hybridization time, the concentration of detergent or solvent, and the inclusion or exclusion of carrier DNA, are well known to those skilled in the art. Additional variations on these conditions will be readily apparent to those skilled in the art (Wahl and Berger (1987) Methods Enzymol 152:399-407; Kimmel (1987) Methods Enzymol 152:507-511; Ausubel supra; and Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview NY).

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NSEQ or the polynucleotide sequences encoding PSEQ can be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to

detect upstream sequences, such as promoters and regulatory elements. (See, e.g.,
Dieffenbach and Dveksler (1995) PCR Primer, a Laboratory Manual, Cold Spring Harbor
Press, Plainview NY; Sarkar (1993) PCR Methods Applic 2:318-322; Triglia et al. (1988)
Nucleic Acids Res 16:8186; Lagerstrom et al. (1991) PCR Methods Applic 1:111-119; and
Parker et al. (1991) Nucleic Acids Res 19:3055-306). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto, CA) to walk genomic
DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 Primer Analysis software (National Biosciences,
Plymouth MN) or another appropriate program, to be about 18 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

In another aspect of the invention, NSEQ or the polynucleotide sequences encoding PSEQ can be cloned in recombinant DNA molecules that direct expression of PSEQ or the polypeptides encoded by NSEQ, or structural or functional fragments thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express the polypeptides of PSEQ or the polypeptides encoded by NSEQ. The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter the nucleotide sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

In order to express a biologically active polypeptide encoded by NSEQ, NSEQ or the polynucleotide sequences encoding PSEQ, or derivatives thereof, may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible

promoters, and 5' and 3' untranslated regions in the vector and in NSEQ or polynucleotide sequences encoding PSEQ. Methods which are well known to those skilled in the art may be used to construct expression vectors containing NSEQ or polynucleotide sequences encoding PSEQ and appropriate transcriptional and translational control elements. These methods include in vitro recombinant DNA techniques, synthetic techniques, and in vivo genetic recombination. (See, e.g., Sambrook (supra) and Ausubel (supra).

A variety of expression vector/host cell systems may be utilized to contain and express NSEQ or polynucleotide sequences encoding PSEQ. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (baculovirus); plant cell systems transformed with viral expression vectors, cauliflower mosaic virus (CaMV) or tobacco mosaic virus (TMV), or with bacterial expression vectors (Ti or pBR322 plasmids); or animal cell systems. The invention is not limited by the host cell employed. For long term production of recombinant proteins in mammalian systems, stable expression of a polypeptide encoded by NSEQ in cell lines is preferred. For example, NSEQ or sequences encoding PSEQ can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector.

In general, host cells that contain NSEQ and that express PSEQ may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences. Immunological methods for detecting and measuring the expression of PSEQ using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS).

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Host cells transformed with NSEQ or polynucleotide sequences encoding PSEQ may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly

PCT/US99/23315 WO 00/21986

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depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of NSEQ or polynucleotides encoding PSEQ may be designed to contain signal sequences which direct secretion of PSEQ or polypeptides encoded by NSEQ through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38), are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant NSEQ or nucleic acid sequences encoding PSEQ are ligated to a heterologous sequence resulting in translation of a fusion protein containing heterologous protein moieties in any of the aforementioned host systems. Such heterologous protein moieties facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, hemagglutinin (HA) and monoclonal antibody epitopes..

In another embodiment, NSEQ or sequences encoding PSEQ are synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers et al. (1980) Nucleic Acids Symp Ser (7) 215-223; Horn et al. (1980) Nucleic Acids Symp Ser (7) 225-232; and Ausubel, <u>supra</u>). Alternatively, PSEQ or a polypeptide sequence encoded by NSEQ itself, or a fragment thereof, may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge et al. (1995) Science 269:202-204). Automated synthesis may be achieved using the ABI 431A Peptide synthesizer (PE Biosystems, Foster City CA). Additionally, PSEQ or the amino acid sequence encoded by NSEQ, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a polypeptide

variant.

In another embodiment, the invention provides a substantially purified polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NO:21, SEQ ID NO:22, SEQ ID NO:23 or fragments thereof.

5 DIAGNOSTICS and THERAPEUTICS

The sequences of the these genes can be used in diagnosis, prognosis, treatment, prevention, and evaluation of therapies for diseases associated with matrix-remodeling, particularly cancer, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis, atherosclerosis, fibrosis, and ulceration. Further, the amino acid sequences encoded by the novel genes are potential therapeutic proteins and targets of anti-cancer therapeutics or for the treatment of other diseases associated with matrix remodeling.

In one preferred embodiment, the polynucleotide sequences of NSEQ or the polynucleotides encoding PSEQ are used for diagnostic purposes to investigate the altered expression of PSEQ, and to monitor regulation of the levels of mRNA or the polypeptides encoded by NSEQ during therapeutic intervention. The polynucleotides may be at least 18 nucleotides long, and may be complementary RNA or DNA molecules, branched nucleic acids, or peptide nucleic acids (PNAs). Alternatively, the polynucleotides are used to detect and quantitate gene expression in samples in which expression of PSEQ or the polypeptides encoded by NSEQ are correlated with disease. Additionally, NSEQ or the polynucleotides encoding PSEQ can be used to detect genetic polymorphisms associated with a disease. These polymorphisms may be detected at the transcript cDNA or genomic level.

The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification (maximal, high, intermediate, or low), will determine whether the probe identifies only naturally occurring sequences encoding PSEQ, allelic variants, or related sequences.

Probes may also be used for the detection of related sequences, and should preferably have at least 70% sequence identity to any of the NSEQ or PSEQ-encoding sequences.

Means for producing specific hybridization probes for DNAs encoding PSEQ include the cloning of NSEQ or polynucleotide sequences encoding PSEQ into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available,

and may be used to synthesize RNA probes in vitro by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin 5 coupling systems, by fluorescent labels and the like. The polynucleotide sequences encoding PSEQ may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; and in microarrays utilizing fluids or tissues from patients to detect altered NSEQ expression. Such qualitative or quantitative methods are well known in the art.

NSEQ or the nucleotide sequences encoding PSEQ can be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantitated and compared with a standard value, typically, derived from a non-diseased sample. If the amount of signal in the patient sample is significantly 15 altered in comparison to the standard value then the presence of altered levels of nucleotide sequences of NSEQ and those encoding PSEQ in the sample indicates the presence of the associated disease. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

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Once the presence of a disease is established and a treatment protocol is initiated, hybridization or amplification assays can be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in a healthy subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

The polynucleotides may be used for the diagnosis of a variety of diseases associated with matrix-remodeling including cancer such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus, cardiomyopathy, arthritis, angiogenesis, diabetic necrosis, atherosclerosis, fibrosis, and ulceration.

Alternatively, the polynucleotides may be used as targets in a microarray. The microarray can be used to monitor the expression level of large numbers of genes simultaneously and to identify splice variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disease, to diagnose a disease, and to develop and monitor the activities of therapeutic agents.

In yet another alternative, polynucleotides may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Fluorescent <u>in situ</u> hybridization (FISH) may be correlated with other physical chromosome mapping techniques and genetic map data. (See, e.g., Heinz-Ulrich et al. (1995) in Meyers, <u>supra</u>, pp. 965-968.)

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In another embodiment, antibodies which specifically bind PSEO may be used for the diagnosis of diseases characterized by the over-or-underexpression of PSEQ or polypeptides encoded by NSEQ. A variety of protocols for measuring PSEQ or the polypeptides encoded by NSEQ, including ELISAs, RIAs, and FACS, are well known in the art and provide a basis for diagnosing altered or abnormal levels of the expression of PSEQ or the polypeptides encoded by NSEQ. Standard values for PSEQ expression are established by combining body fluids or cell extracts taken from healthy subjects, preferably human, with antibody to PSEO or a polypeptide encoded by NSEQ under conditions suitable for complex formation. The amount of complex formation may be quantitated by various methods, preferably by photometric means. Quantities of PSEQ or the polypeptides encoded by NSEQ expressed in disease samples from, for example, biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing or monitoring disease. Alternatively, one may use competitive drug screening assays in which neutralizing antibodies capable of binding PSEQ or the polypeptides encoded by NSEQ specifically compete with a test compound for binding the polypeptides. Antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with PSEQ or the polypeptides encoded by NSEQ.

In another aspect, the polynucleotides and polypeptides of the present invention can be employed for treatment or the monitoring of therapeutic treatments for cancer. The polynucleotides of NSEQ or those encoding PSEQ, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, the complement of the polynucleotides of NSEQ or those encoding PSEQ may be used in situations in which it would be desirable to

block the transcription or translation of the mRNA.

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Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. Methods which are well known to those 5 skilled in the art can be used to construct vectors to express nucleic acid sequences complementary to the polynucleotides encoding PSEQ. (See, e.g., Sambrook, supra; and Ausubel, supra.)

Genes having polynucleotide sequences of NSEQ or those encoding PSEQ can be turned off by transforming a cell or tissue with expression vectors which express high levels of 10 a polynucleotide, or fragment thereof, encoding PSEQ. Such constructs may be used to introduce untranslatable sense or antisense sequences into a cell. Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, are preferred. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee et al. (1994) In: Huber and Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. Alternatively, nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and 25 similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases may be included.

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back 30 into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g.,

Goldman et al. (1997) Nature Biotechnology 15:462-466.)

Further, an antagonist or antibody of a polypeptide of PSEQ or encoded by NSEQ may be administered to a subject to treat or prevent a cancer associated with increased expression or activity of PSEQ. An antibody which specifically binds the polypeptide may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissue which express the the polypeptide.

Antibodies to PSEQ or polypeptides encoded by NSEQ may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are especially preferred for therapeutic use. Monoclonal antibodies to PSEQ may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. In addition, techniques developed for the production of chimeric antibodies can be used. (See, for example, Meyers, supra.) Alternatively, techniques described for the production of single chain antibodies may be employed. Antibody fragments which contain specific binding sites for PSEQ or the polypeptide sequences encoded by NSEQ may also be generated.

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art.

Yet further, an agonist of a polypeptide of PSEQ or that encoded by NSEQ may be administered to a subject to treat or prevent a cancer associated with decreased expression or activity of the polypeptide.

An additional aspect of the invention relates to the administration of a pharmaceutical or sterile composition, in conjunction with a pharmaceutically acceptable carrier, for any of the therapeutic effects discussed above. Such pharmaceutical compositions may consist of polypeptides of PSEQ or those encoded by NSEQ, antibodies to the polypeptides, and mimetics, agonists, antagonists, or inhibitors of the polypeptides. The compositions may be administered alone or in combination with at least one other agent, such as a stabilizing

PCT/US99/23315 WO 00/21986

compound, which may be administered in any sterile, biocompatible pharmaceutical carrier including, but not limited to, saline, buffered saline, dextrose, and water. The compositions may be administered to a patient alone, or in combination with other agents, drugs, or hormones.

The pharmaceutical compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

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In addition to the active ingredients, these pharmaceutical compositions may contain suitable pharmaceutically-acceptable carriers comprising excipients and auxiliaries which facilitate processing of the active compounds into preparations which can be used pharmaceutically. Further details on techniques for formulation and administration may be found in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells or in animal models such as mice, rats, rabbits, dogs, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example, polypeptides of PSEQ or those encoded by NSEQ, or fragments thereof, antibodies of the polypeptides, and agonists, antagonists or inhibitors of the polypeptides, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED_{50} (the dose therapeutically effective in 50% of the population) or LD_{50} (the dose lethal to 50% of the population) statistics.

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as dogs, cats, cows, horses, rabbits, monkeys, and most preferably, humans.

EXAMPLES

It is understood that this invention is not limited to the particular methodology,

protocols, and reagents described, as these may vary. It is also understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims. The examples below are provide to illustrate the subject invention and are not included for the purpose of limiting the invention.

I cDNA Library Construction

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The cDNA library, THYMFET02, was selected to demonstrate the construction of the cDNA libraries from which novel matrix remodeling genes were derived. The THYMFET02 cDNA library was constructed from microscopically normal thymus tissue obtained from a Caucasian female fetus who died at 17 weeks gestation from anencephaly. Serology was negative; family history included tobacco abuse and gastritis.

The frozen tissue was homogenized and lysed in TRIZOL reagent (1 gm tissue/10 ml TRIZOL; Life Technologies, Rockville MD), a monoplastic solution of phenol and guanidine isothiocyanate, using a POLYTRON homogenizer (PT-3000; Brinkmann Instruments, Westbury NY). After a brief incubation on ice, chloroform was added (1:5 v/v), and the lysate

was centrifuged. The upper chloroform layer was removed, and the RNA was precipitated with isopropanol, resuspended in DEPC-treated water, and treated with DNase for 25 min at 37°C. The mRNA was reextracted once with acid phenol-chloroform pH 4.7 and precipitated using 0.3 M sodium acetate and 2.5 volumes ethanol. The mRNA was isolated using the OLIGOTEX kit (Qiagen, Chatsworth CA) and used to construct the cDNA library.

The mRNA was handled according to the recommended protocols in the SUPERSCRIPT Plasmid system (Life Technologies). The cDNAs were fractionated on a SEPHAROSE CL4B column (Amersham Pharmacia Biotech, Pisctaway NJ), and those cDNAs exceeding 400 bp were ligated into pINCY 1 plasmid (Incyte Pharmaceuticals, Palo Alto CA). The plasmid was subsequently transformed into DH5α competent cells (Life Technologies).

II Isolation and Sequencing of cDNA Clones

Plasmid DNA was released from the cells and purified using the REAL Prep 96

Plasmid kit (Qiagen). This kit enabled the simultaneous purification of 96 samples in a 96well block using multi-channel reagent dispensers. The recommended protocol was employed except for the following changes: 1) the bacteria were cultured in 1 ml of sterile Terrific Broth

(Life Technologies) with carbenicillin at 25 mg/L and glycerol at 0.4%; 2) after inoculation, the cultures were incubated for 19 hours and at the end of incubation, the cells were lysed with 0.3 ml of lysis buffer; and 3) following isopropanol precipitation, the plasmid DNA pellet was resuspended in 0.1 ml of distilled water. After the last step in the protocol, samples were transferred to a 96-well block for storage at 4°C.

The cDNAs were prepared using a MICROLAB 2200 (Hamilton, Reno NV) in combination with DNA ENGINE thermal cyclers (PTC200; MJ Research, Watertown MA) and sequenced by the method of Sanger et al. (1975, J Mol Biol 94:441f) using ABI PRISM 377 DNA sequencing systems.

Selection, Assembly, and Characterization of Sequences 10 III

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The sequences used for coexpression analysis were assembled from EST sequences, 5' and 3' longread sequences, and full length coding sequences. Selected assembled sequences were expressed in at least three cDNA libraries.

The assembly process is described as follows. EST sequence chromatograms were processed and verified. Quality scores were obtained using PHRED (Ewing et al. (1998) Genome Res 8:175-185; Ewing and Green (1998) Genome Res 8:186-194). Then the edited sequences were loaded into a relational database management system (RDBMS). The EST sequences were clustered into an initial set of bins using BLAST with a product score of 50. All clusters of two or more sequences were created as bins. The overlapping sequences 20 represented in a bin correspond to the sequence of a transcribed gene.

Assembly of the component sequences within each bin was performed using a modification of PHRAP, a publicly available program for assembling DNA fragments (Phil Green, University of Washington, Seattle WA). Bins that showed 82% identity from a local pair-wise alignment between any of the consensus sequences were merged.

Bins were annotated by screening the consensus sequence in each bin against public databases, such as GBpri and GenPept from NCBI. The annotation process involved a FASTn screen against the GBpri database in GenBank. Those hits with a percent identity of greater than or equal to 70% and an alignment length of greater than or equal to 100 base pairs were recorded as homolog hits. The residual unannotated sequences were screened by FASTx 30 against GenPept. Those hits with an E value of less than or equal to 10⁻⁸ are recorded as homolog hits.

Sequences were then reclustered using BLASTn and Cross-Match, a program for rapid protein and nucleic acid sequence comparison and database search (Green, <u>supra</u>), sequentially. Any BLAST alignment between a sequence and a consensus sequence with a score greater than 150 was realigned using cross-match. The sequence was added to the bin whose consensus sequence gave the highest Smith-Waterman score amongst local alignments with at least 82% identity. Non-matching sequences created new bins. The assembly and consensus generation processes were performed for the new bins.

IV Coexpression Analyses of Known Matrix-remodeling Genes

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Twenty one known matrix-remodeling genes were selected to identify novel genes that are closely associated with matrix remodeling. The known genes were osteonectin (BM-40), chondroitin/dermatan sulfate proteoglycans (C/DSPG), collagen I, II, II, and IV (coll-I, coll-II, and coll-III), connective tissue growth factor (CTGF), fibrillin, fibronectins, fibronectin receptor (fibr-r), fibulin 1, heparan sulfate proteoglycans (HSPG), extracellular matrix protein (hevin), insulin-like growth factor 1 (IGF 1), insulin-like growth factor binding protein (IGFBP), laminin, lumican, matrix Gla protein (MGP), matrix metalloproteases (MMP), and tissue inhibitors of matrix metalloproteinase 1, 2, and 3 (TIMP 1, 2, and 3). The protein products of the known matrix-remodeling genes may be categorized as follows.

- 1. Extracellular matrix component protein. These proteins include collagens, proteoglycans, fibrillin, fibronectin, fibulin, and laminin that constitute the major structures of the extracellular matrix.
- 2. Matrix proteases and matrix protease inhibitors. These proteins include matrix metalloproteases (MMPs) such as the collagenases, and MMP inhibitors such as the tissue-inhibitors of matrix metalloproteases (TIMPs).
- 3. Regulatory proteins that control expression of matrix-remodeling genes. Such regulatory proteins include connective tissue growth factor, insulin-like growth factor, osteonectin (BM-40), and the receptors for and inhibitors of these proteins.

The known matrix-remodeling genes that we examined in this analysis, and brief descriptions of their functions, are listed in Table 4. Detailed descriptions of their roles in matrix remodeling may be found in the cited articles and reviews.

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Table 4. Known Matrix-remodeling Genes.

Gene	Description & References
BM-40	Alternate names: SPARC, osteonectin
	Regulates connective tissue remodeling, wound healing, angiogenesis
	Induces matrix metalloprotease synthesis (collagenase & gelatinase)
	Regulates cell movement and proliferation
	Expression increased in neoplastic melanoma, fibrosis, angiogenesis. (Kamihagi et al. (1994) Biochem Biophys Res Commun 200:423-8; Lane et al. (1994) J Cell Biol 125:929-43; Inagaki et al. (1996) Life Sci 58:927-34; Ledda et al. (1997) J Invest Dermatol 108:210-4; Shankavaram et al. (1997) J Cell Physiol 173:327-34.)
C/DSPG	Chondroitin/dermatan sulfate proteoglycans
	Major extracellular matrix proteoglycan
	Regulate cell proliferation, attachment and migration. Darnell et al. (1990) Molecular Cell Biology, Scientific American Press, New York NY; Toole (1991) In: Cell Biology of Extracellular Matrix, Plenum, New York NY, pp. 305-341; Beck et al. (1993) Biochem Biophys Res Commun 190:616-23)
Collagens	Family of fibrous structural proteins (collagen I, II, III, IV, etc.)
	Most abundant structural component of the extracellular matrix Secreted as procollagen; converted to collagen by MMPs
	(Alexander and Werb (1991) In: <u>Cell Biology of Extracellular Matrix</u> , pp. 255-302; Adams (1993) In: <u>Extracellular Matrix</u> , Marcel Dekker, New York, NY pp. 91-119; Schuppan et al. (1993) In: <u>Extracellular Matrix</u> , pp. 201-254.)
CTGF	Connective tissue growth factor
	Mediates induction of matrix synthesis and fibrosis (Grotendorst (1997) Cytokine Growth Factor Rev 8:171-9; Oemar and Luscher (1997) Arterioscler Thromb Vasc Biol 17:1483-9; Ito et al. (1998) Kidney Int 53:853-61.)
fibrillin	Major component of extracellular microfibrills (matrix elastic network)
THO THIN	Present in connective tissue throughout the body (Kielty and Shuttleworth (1995) Int J Biochem Cell Biol 27:747-60; Haynes et al. (1997) Br J Dermatol 137:17-23; Hayward and Brock (1997) Hum Mutat 10:415-23.)
fibronectins	Family of extracellular matrix glycoproteins Anchor cells to the matrix
	Bind matrix proteins to cell surface receptors
fibr-r	Fibronectin receptor Fibronectin receptors regulate cell adhesion & migration (Darnell et al. (1990) Molecular Cell Biology, Scientific American Press, New York NY; Ruoslahti (1991) Cell Biology of Extracellular Matrix, pp. 343-363; Yamada (1991) Cell Biology of Extracellular Matrix, pp. 111-146.)
fibulin 1	Fibronectin-binding extracellular matrix protein

MGP

		Mediates platelet adhesion via a bridge of fibrinogen Cleaved by matrix metalloproteinases Inhibits breast and ovarian cancer cell motility (Argraves et al. (1990) J Cell Biol 111:3155-64; Sasaki et al. (1996) Eur J Biochem 240:427-34; Hayashido et al. (1998) Int J Cancer 75:654-8.)
5	HSPG	Heparan sulfate proteoglycans Extracellular matrix proteoglycan found on cell surface of many cell types Regulate cell interactions with the extracellular matrix Bind to collagens and fibronectin in the matrix
10		Regulate cell proliferation, attachment and migration (Darnell et al. (1990); Toole (1991) In: Cell Biology of Extracellular Matrix, pp. 305-341; Schuppan et al. (1993) In: Extracellular Matrix, pp. 201-254.)
15	hevin	Extracellular matrix protein Homolog to BM-40 Regulates cell adhesion and migration Downregulated in metastatic prostate cancer, lung cancer (Girard and Springer (1996) J Biol Chem 271:4511-7; Bendik et al. Cancer Res 58:232-6.)
20	IGF 1	Insulin-like growth factor Regulates matrix homeostatis and remodeling Regulates aggregation, growth and survival of cancer cells (Aston et al. (1995) Am J Respir Crit Care Med 151:1597-603; Bitar and Labbad (1996) J Surg Res 61:113-9; Guvakova and Surmacz (1997) Exp Cell Res 231:149-62; Sunic et al. (1998) Endocrinology 139:2356-62.)
25	IGFBP	Insulin-like growth factor binding protein Regulates IGF-1 bioavailability (binds IGF-1 more strongly than the receptor) Degraded by matrix metalloproteases (Kiefer et al. (1991) Biochem Biophys Res Commun 176:219-25; Fowlkes et al. (1995) Prog Growth Factor Res 6:255-63; Parker et al. (1996) J Biol Chem 271:13523-9.)
30	laminin	Major protein in basal lamina, with collagen, HSPG, and entactin Anchors cells to the matrix by binding collagen, HSGP and heparin Laminins and collagens are the main targets of MMPs Regulates cell attachment, migration, growth, and differentiation (Yamada et al. (1993) In: Extracellular Matrix, pp. 49-66; Giannelli et al. (1997) Science 277:225-8; Quaranta and Plopper (1997) Kidney Int 51: 1441-6; Soini et al. (1997) Hum Pathol 28:220-6.)
35	lumican	Extracellular proteoglycan Organizes collagen fibrils in extracellular matrix (Dourado et al. (1996) Osteoarthritis Cartilage 4:187-96; Scott (1996) Biochemistry 35:8795-9; Cs-Szabo et al. (1997) Arthritis Rheum 40:1037-45.)

Matrix Gla protein Regulates calcification of cartilage

Marker for osteoblast activity (Shanahan et al. (1994) J Clin Invest 93:2393-402; Luo et al. (1997) Nature 386:78-81; Martinetti et al. (1997) Tumour Biol 18:197-205)

MMP Family of Matrix Metalloproteases (including collagenases)

Cleave procollagen to produce collagen

(Alexander and Werb (1991) In: Cell Biology of Extracellular Matrix, pp. 255-302; Adams (1993) In: Extracellular Matrix, pp. 91-119; Schuppan et al. (1993) In: Extracellular Matrix pp. 201-254.)

TIMP 1, 2, 3 Tissue inhibitors of matrix metalloproteinases
Bind and inactivate matrix proteases
(Schuppan et al. (1993) In: Extracellular Matrix, pp. 201-254; Zvibel and Kraft (1993) In: Extracellular Matrix, pp. 559-580.)

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The coexpression of the 21 known genes with each other is shown in Table 5. The entries in Table 5 are the negative log of the p-value $(-\log p)$ for the coexpression of the two genes. As shown, the method successfully identified the strong association of the known genes among themselves, indicating that the coexpression analysis method of the present invention was effective in identifying genes that are closely associated with matrix remodeling.

Table 5. Coexpression of 21 known matrix-remodeling genes. $(-\log p)$

	_																				
	laminin	fibrillin	lumican	≥ ≣∞	TIMP-1	IGFBP	IN 1100	TIMP-3	CTGF	hevin	fibulin	BM-40	TIMP-2	HSPG	fibronec	MGP	C/DSP	fibr-r	1-lloo	III-iloo	MMP
	laminin	7	9	21	9	15	8	4	5	7	14	10	7	11	9	19	11	7	16	10	13
	fibrillin 7		13	8	6	7	14	11	4	7	12	7	8	4	8	6	13	6	11	12	11
	lumican 9	13		24	17	16	28	17	17	14	15	22	10	8	12	25	33	14	32	34	17
25	coll IV 21	8	24		17	22	22	13	11	14	28	25	12	22	16	27	26	12	34	25	26
	TIMP-1 9	6	17	17		20	15	11	11	6	10	21	15	9	16	20	13	8	14	20	19
	IGFBP 15	7	16	22	20		20	18	16	11	14	18	14	19	21	25	23	10	27	23	20
	coll VI 8	14	28	22	15	20		13	17	19	16	20	11	11	19	19	28	12	31	36	27
	TIMP-3 4	11	17	13	11	18	13		13	18	20	22	14	9	10	18	25	12	12	13	9
30	CTGF 5	4	17	11	11	16	17	13		8	10	18	7	7	19	22	12	12	18	13	11
	hevin 7	. 7	14	14	6	11	19	18	8		15	18	13	8	8	23	27	10	14	11	8
	fibulin 14	12	15	28	10	14	16	20	10	15		19	9	11	8	19	20	6	17	20	18
	BM-40 10	7	22	25	21	18	20	22	18	18	19		14	11	24	21	24	16	25	32	19
	TIMP-2 7	8	10	12	15	14	11	14	7	13	9	14		7	12	8	16	11	13	13	13
35	HSPG 11	4	8	22	9	19	11	9	7	8	11	11	7	_	8	14	10	6	11	10	10
	fibronec 9	8	12	16	16	21	19	10	19	8	8	24	12	8		14	14	11	24	21	15
	MGP 19	6	25	27	20	25	19	18	22	23	19	21	8	14	14		32	14	25	20	13
	C/DSPG 11	13	33	26	13	23	28	25	12	27	20	24	16	10	14	32		14	27	28	14
40	fibr-r 7	6	14	12	. 8	10	12	12	12	10	6	16	11	6	11	14	14		14	13	6
40	coll-I 16	11	32	34	14	27	31	12	18	14	17	25	13	11	24	25	27	14		42	21
	coll-III 10	12	34	25	20	23	36	13	13	11	20	32	13	10	21	20	28	13	42		23
	<u>MMP 13</u>	11_	17	26	19	20	<u>27</u>	9	11_	8	18	19_	13	10	<u> 15</u>	<u>13</u>	14	_6_	21	<u>23</u>	

45 V Novel Genes Associated with Matrix Remodeling

Using coexpression analysis, we have identified 20 novel genes that show strong association with known matrix remodeling genes from a total of 41,419 assembled gene

sequences. The degree of association was measured by probability values and has a cutoff of p value less than 0.00001. This was followed by annotation and literature searches to insure that the genes that passed the probability test have strong association with known matrix-remodeling genes. This process was reiterated so that the initial 41,419 genes were reduced to the final 20 matrix-remodeling genes. Details of the coexpression patterns for the 20 novel matrix-remodeling genes are presented in Table 6.

Each of the 20 novel genes is coexpressed with at least two of the 21 known genes with a p-value of less than 10⁻⁷. The coexpression results are shown in Table 6.

The novel genes identified are listed in the table by their Incyte clone numbers (Clone), and the known genes their abbreviated names (Gene) as shown in Example IV.

Table 6. Coexpression of 20 novel genes with known matrix-remodeling genes. (- $\log p$)

15	Gene	laminin	fibrillin	lumican	≥ 00	TIMP-1	IGFBP	 	TIMP-3	CTGF	hevin	fibulin	BM-40	TIMP-2	HSPG	fibronec	MGP	C/DSP	fibr-r	8	=- 8	MMP
•	606132	8	7	2	6	4	7	7	2	4	4	4	3	3	4	4	3	2	2	5	3	10
	627722	3	4	1	1	3	3	2	5	3	6	3	4	3	2	6	5	3	3	2	3	4
	639644	6	7	11	10	3	4	7	3	14	6	6	9	6	2	9	8	5	6	9	7	6
	1362659	6	5	6	7	6	9	10	9	8	8	7	6	8	6	7	9	9	7	10	5	5
20	1446685	6	6	11	13	4	7	8	5	7	5	10	9	5	9	5	9	8	6	8	10	7
	1556751	3	7	7	8	8	9	9	8	7	6	5	5	7	8	4	10	11	3	7	6	8
	1656953	6	8	6	2	5	7	8	5	6	9	3	7	4	3	4	10	8	7	4	4	5
	1662318	9	3	6	10	7	9	5	5	8	8	6	8	5	9	6	8	6	4	7	7	9
	1996726	3	4	7	7	6	5	8	3	10	2	2	3	2	2	9	3	6	6	8	11	6
25	2137155	3	2	6	3	4	2	2	4	6	4	2	9	4	2	8	4	4	4	5	2	5
	2268890	9	13	7	9	8	11	8	9	5	5	8	7	8	5	8	8	11	3	11	7	11
	2305981	3	2	4	6	3	4	3	5	5	6	7	5	2	2	2	7	6	4	3	2	2
	2457612	3	3	3	5	2	4	4	2	8	4	5	5	2	2	7	8	6	6	5	4	8
	2814981	6	3	5	7	4	6	7	2	2	5	5	5	3	6	5	4	6	1	6	4	7
30	3089150	4	6	11	8	5	10	13	9	14	10	11	10	7	6	8	11	16	11	9	7	5
	3206667	8	5	10	9	7	5	6	4	9	4	7	8	4	4	7	13	12	4	8	8	6
	3284695	7	6	7	14	8	7	6	14	8	18	12	9	10	8	6	18	10	5	13	6	6
	3481610	3	2	4	4	3	6	4	6	6	7	4	5	1	5	5	7	5	3	3	2	2
	3722004	6	4	8	10	13	9	7	13	8	9	11	12	11	5	10	9	12	3	7	7	6
35.	3948614	11	8	_6	17	8	13	12	5_	5_	11	12	<u> 7</u>	11	13	4	_ 7_	_7_	_4	14	11	10

VI Novel Genes Associated with Matrix Remodeling

The 20 novel genes were identified from the data shown in Table 6 to be associated with matrix remodeling.

The nucleotide sequences comprising the consensus sequences of SEQ ID NOs:1-20 of the present invention were first identified from Incyte Clones 606132, 627722, 639644, 1362659, 1446685, 1556751, 1656953, 1662318, 1996726, 2137155, 2268890, 2305981, 2457612, 2814981, 3089150, 3206667, 3284695, 3481610, 3722004, and 3948614, respectively, and assembled according to Example III. BLAST and other motif searches were performed for SEQ ID NOs:1-20 according to Example VII. The sequences of SEQ ID NOs:1-20 were translated and sequence identity was sought with known sequences. Polypeptide sequences comprising the consensus sequences of SEQ ID NO:21, SEQ ID NO:22, and SEQ ID NO:23 of the present invention were encoded by SEQ ID NO:2,

PCT/US99/23315 WO 00/21986

SEQ ID NO:6, and SEQ ID NO:11, respectively. SEQ ID NOs:21-23 were analyzed using BLAST and other motif search tools as disclosed in Example VII.

SEQ ID NO:3 is 2987 residues in length and shows about 59% sequence identity from about nucleotide 2117 to about nucleotide 2914 with the cDNA encoding regulatory subunit of a human cAMP-dependent protein kinase, RIIbeta (WO 88/03164). SEQ ID NO:8 is 3017 nucleotides in length and shows about 70% to about 74% sequence identity from about nucleotide 1 to about nucleotide 1260 and about nucleotide 1925 to about nucleotide 1985 with human Hpast mRN (g2529706), a gene associated with multiple endocrine neoplasia type 1. SEQ ID NO:9 is 1735 nucleotides in length and shows about 25% sequence identity from about nucleotide 5 to about nucleotide 1534 with a human neuronal cell adhesion molecule (WO 96/04396) important in the development of nervous system by promoting cell-cell adhesion. SEQ ID NO:14 is 2040 nucleotides in length and shows about 60% to 70% sequence identity from about nucleotide 1 to about nucleotide 1023 with a human mRNA for a serine protease (g1621243) specific for insulin-like growth factor-binding proteins. The amino acid sequence encoded by SEQ ID NO:14 from about nucleotide 3 to about nucleotide 1043 shows about 61% sequence identity with an osteoblast-like cell-derived protein (J09107980) useful for treatment and prevention of various diseases and as contraceptive. SEQ ID NO:15 is 2121 nucleotides in length and shows 60-80% sequence identity with a mouse gene, ADAMT-1 (g2809056), a member of the ADAM (the disintegrin and metalloproteinase) family. ADAMT-1 has been shown to contain the thrombospondin (TSP) type I motif; expression of ADAMT-1 is closely associated with inflammatory processes (Kuno et al (1997) Genomics 46:466-471). SEQ ID NO:16 is 2900 nucleotides in length and shows about 70% sequence identity with a mouse homeobox (Pmx) mRNA (g460124). Homeobox genes are expressed in very specific temporal and spatial pattern and function as transcriptional regulators of developmental processes (Kern et al. (1994) Genomics 19:334-340).

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SEQ ID NO:21 is 551 amino acid residues long and shows about 37% sequence identity from about amino acid residue 10 to about amino acid residue 278 with PALM (g3219602), a human paralemin that is membrane-bound and expressed abundantly in brain and at intermediate levels in the kidney and in endocrine cells. In addition, the sequence encompassing residues 418 to 434 of SEQ ID NO:21 resembles one of the structural fingerprint regions of a seven trans-membrane receptor, LCR1. that is isolated from the human brain (Rimland et al. (1991) Mol Pharmacol 40:869-875). SEO ID 30 NO:21 also has one potential amidation site at L546; three potential N-glycosylation sites at N223, N229, and N408; one potential cAMP- and cGMP-dependent protein kinase phosphorylation site at S486; fifteen potential casein kinase II phosphorylation sites at S57, S100, T101, T116, S135, S253, T349, S370, T387, S426, T434, S489, S505, S520, and T526; one potential N-myristoylation site at G54; and nine potential protein kinase C phosphorylation sites at T15, S25, S57, S100, S123, S247.

S364, S370, and S505. SEQ ID NO:22 is 99 amino acid residues in length. The sequence of SEQ ID NO:22 from about amino acid residue 71 to about amino acid residue 81 resembles one of the fingerprint regions of the RH1 and RH2 opsins, a family of G protein coupled receptors that mediate vision (Zuker et al. (1985) Cell 40:851-858; Cowman et al. (1986) Cell 44:705-710). SEQ ID NO:22 also has one potential N-myristoylation site at G24, and two potential protein kinase C phosphorylation sites at S13 and S89. SEQ ID NO:23 is 493 amino acid residues in length and shows about 44% sequence identity from about amino acid residue 277 to about amino acid residue 487 with an angiopoietin-like factor from the human cornea, CDT6 (g2765527). Angiopoietin 1 and angiopoietin 2 function as a natural ligand and a natural inhibitor, respectively, for TIE2, a receptor 10 critical in angiogenesis during embryonic development, tumor growth, and tumor metastasis. The sequences encompassing amino acid residues 305 to 343, 346 to 355, 365 to 402, 411 to 424, and 428 to 458 of SEQ ID NO:23 resemble the carboxy-terminal domain signatures of fibrinogen beta and gamma chains from BLOCKS analysis. SEQ ID NO:23 also exhibits one potential signal peptide region encompassing amino acid residues M1 to G22 when analyzed using a HMM-based signal peptide analysis tool. In addition, SEQ ID NO:23 shows two potential N-glycosylation sites at N164 and N192; one potential cAMP- and cGMP-dependent protein kinase phosphorylation sites at S127, six potential casein kinase II phosphorylation sites at S34, S209, T238, S266, T368, and T417; four potential N-myristoylation sites at G12, G18, G22, and G29; eight potential protein kinase C phosphorylation sites at S34, S209, T268, T299, T335, S373, S383, and S477; and three potential tyrosine kinase phosphorylation sites at Y183, Y392, and Y467. 20

VII Homology Searching for Matrix-Remodeling Renes and the Proteins Encoded by the Genes

Polynucleotide sequences, SEQ ID NOs:1-20, and polypeptide sequences, SEQ ID NOs: 21-23, were queried against databases derived from sources such as GenBank and SwissProt. These databases, which contain previously identified and annotated sequences, were searched for regions of similarity using Basic Local Alignment Search Tool (BLAST; Altschul (1990) supra) and Smith-Waterman alignment (Smith et al. (1992) Protein Engineering 5:35-51). BLAST searched for matches and reported only those that satisfied the probability thresholds of 10-25 or less for nucleotide sequences and 10-8 or less for polypeptide sequences.

The polypeptide sequences were also analyzed for known motif patterns using MOTIFS, SPSCAN, BLIMPS, and Hidden Markov Model (HMM)-based protocols. MOTIFS (Genetics Computer Group, Madison WI) searches polypeptide sequences for patterns that match those defined in the Prosite Dictionary of Protein Sites and Patterns (Bairoch et al. supra), and displays the patterns found and their corresponding literature abstracts. SPSCAN (Genetics Computer Group) searches for

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potential signal peptide sequences using a weighted matrix method (Nielsen et al. (1997) Prot Eng 10:1-6). Hits with a score of 5 or greater were considered. BLIMPS uses a weighted matrix analysis algorithm to search for sequence similarity between the polypeptide sequences and those contained in BLOCKS, a database consisting of short amino acid segments, or blocks, of 3-60 amino acids in length, compiled from the PROSITE database (Henikoff et al. supra; Bairoch et al. supra), and those in PRINTS, a protein fingerprint database based on non-redundant sequences obtained from sources such as SwissProt, GenBank, PIR, and NRL-3D (Attwood et al. (1997) J Chem Inf Comput Sci 37:417-424). For the purposes of the present invention, the BLIMPS searches reported matches with a cutoff score of 1000 or greater and a cutoff probability value of 1.0 x 10⁻³. HMM-based protocols were based on a probabilistic approach and searched for consensus primary structures of gene families in the protein sequences (Eddy, supra; Sonnhammer, supra). More than 500 known protein families with cutoff scores ranging from 10 to 50 bits were selected for use in this invention.

VIII Labeling and Use of Individual Hybridization Probes

Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of [γ - 32 P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (NEN Life Science Products, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine resin column (Amersham Pharmacia Biotech). An aliquot containing 10^7 counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (NEN Life Science Products).

The DNA from each digest is fractionated on a 0.7 percent agarose gel and transferred to nylon membranes (NYTRAN PLUS, Schleicher & Schuell, Durham NH). Hybridization is carried out under the following conditions: 5x SCC/0.1% SDS at 60° C for about 6 hours, subsequent washes are performed at higher stringency with buffers, such as 1x SCC/0.1% SDS at 45° C, then 0.1xSCC. After XOMAT AR film (Eastman Kodak, Rochester NY) is exposed to the blots for several hours, hybridization patterns are compared.

IX Production of Specific Antibodies

SEQ ID NO:20, 21, or 23 substantially purified using polyacrylamide gel electrophoresis (Harrington (1990) Methods Enzymol 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the amino acid sequence is analyzed using LASERGENE software (DNASTAR, Madison WI) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for

selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. Typically, oligopeptides 15 residues in length are synthesized using an ABI 431A peptide synthesizer (PE Biosystems) using Fmoc-chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester to increase

5 immunogenicity. Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide activity by, for example, binding the peptide to plastic, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

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1, 2, and 3.

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What is claimed is:

- A substantially purified polynucleotide comprising a gene that is coexpressed with one or
 more known matrix-remodeling genes in a plurality of biological samples, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor, fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycan, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin, lumican, matrix Gla protein,
 matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase 1, 2, and 3.
 - 2. The polynucleotide of claim 1, comprising a polynucleotide sequence selected from the group consisting of:
 - (a) a polynucleotide sequence selected from the group consisting of SEQ ID NOs:1-20;
- (b) a polynucleotide sequence which encodes the polypeptide sequence of SEQ ID NO: 21, 15 22, or 23;
 - (c) a polynucleotide sequence having at least 70% identity to the polynucleotide sequence of (a) or (b);
 - (d) a polynucleotide sequence comprising at least 18 sequential nucleotides of the polynucleotide sequence of (a), (b), or (c);
- 20 (e) a polynucleotide which hybridizes under stringent conditions to the polynucleotide of (a), (b), (c), or (d); and
 - (f) a polynucleotide sequence which is complementary to the polynucleotide sequence of (a), (b), (c), (d), or (e).
 - 3. A substantially purified polypeptide comprising the gene product of a gene that is coexpressed with one or more known matrix-remodeling genes in a plurality of biological samples, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor, fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycans, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin,
 0 lumican, matrix Gla protein, matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase
 - 4. The polypeptide of claim 3, comprising a polypeptide sequence selected from the group consisting of:
 - (a) the polypeptide sequence of SEQ ID NO:21, 22, or 23;
 - (b) a polypeptide sequence having at least 85% identity to the polypeptide sequence of (a); and

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- (c) a polypeptide sequence comprising at least 6 sequential amino acids of the polypeptide sequence of (a) or (b).
 - 5. An expression vector comprising the polynucleotide of claim 2.
 - 6. A host cell comprising the expression vector of claim 5.
- 7. A pharmaceutical composition comprising the polynucleotide of claim 2 or the polypeptide of claim 3 in conjunction with a suitable pharmaceutical carrier.
 - 8. An antibody which specifically binds to the polypeptide of claim 4.
- 9. A method for diagnosing a disease or condition associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor, fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycans, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin, lumican, matrix Gla protein, matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase 1, 2, and 3, the method comprising the steps of:
 - (a) providing a sample comprising one of more of said coexpressed genes;
 - (b) hybridizing the polynucleotide of claim 2(F) to said coexpressed genes under conditions effective to form one or more hybridization complexes; and
 - (c) detecting the hybridization complexes, wherein the altered level of hybridization complexes compared with the level of hybridization complexes of a nondiseased sample correlates with the presence of the disease or condition.
 - 10. A method for treating or preventing a disease associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a subject in need, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor, fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycans, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin, lumican, matrix Gla protein, matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase 1, 2, and 3, the method comprising the step of administering to said subject in need the pharmaceutical composition of claim 7 in an amount effective for treating or preventing said disease.
 - 11. A method for treating or preventing a disease associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a subject in need, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor,

fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycans, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin, lumican, matrix Gla protein, matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase 1, 2, and 3, the method comprising the step of administering to said subject in need the antibody of claim 8 in an amount effective for treating or preventing said disease.

12. A method for treating or preventing a disease associated with the altered expression of a gene that is coexpressed with one or more known matrix-remodeling genes in a subject in need, wherein each known matrix-remodeling gene is selected from the group consisting of osteonectin, chondroitin/dermatan sulfate proteoglycans, collagen I, II, II, and IV, connective tissue growth factor, fibrillin, fibronectins, fibronectin receptor, fibulin 1, heparan sulfate proteoglycans, extracellular matrix protein, insulin-like growth factor 1, insulin-like growth factor binding protein, laminin, lumican, matrix Gla protein, matrix metalloproteases, and tissue inhibitors of matrix metalloproteinase 1, 2, and 3, the method comprising the step of administering to said subject in need the polynucleotide sequence of claim 2(F) in an amount effective for treating or preventing said disease.

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Trp Thr Val Ile Gln Arg Arg Leu Asp Gly Ser Val Asn Phe Phe

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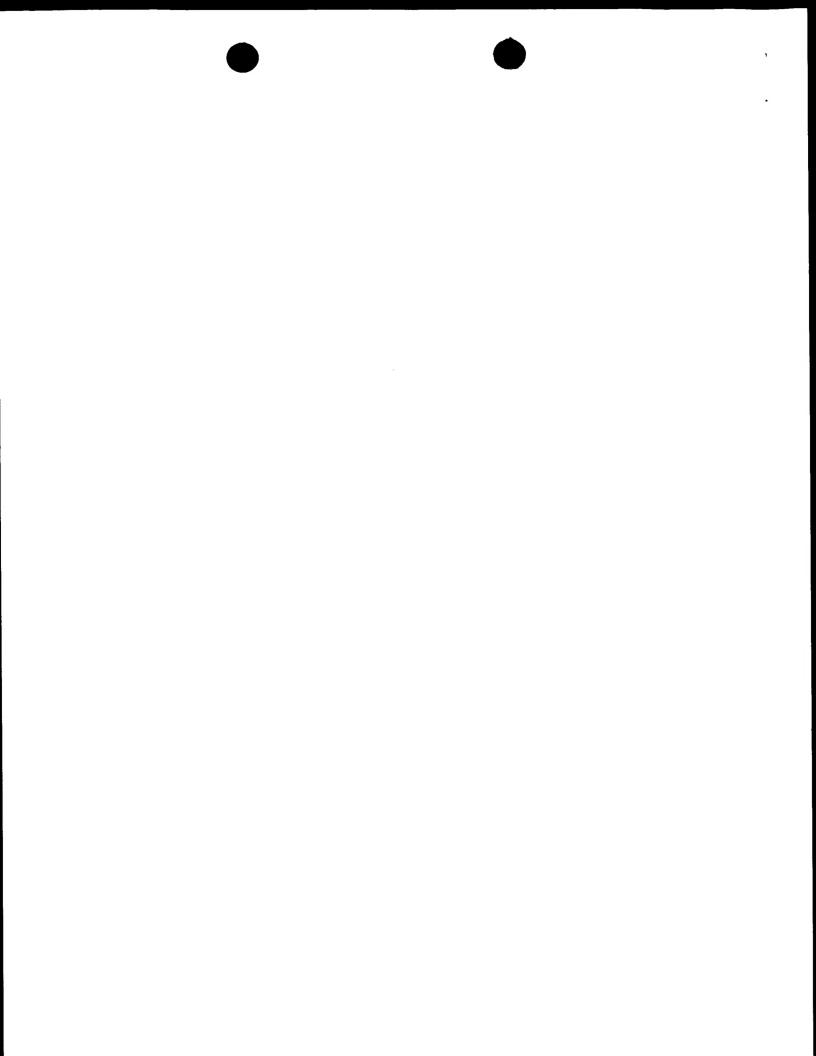
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PCT/US99/23315

WO 00/21986

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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 7: WO 00/21986 (11) International Publication Number: **A3** C12N 9/00, C07K 14/00 (43) International Publication Date: 20 April 2000 (20.04.00) (81) Designated States: AL, AM, AT, AU, AZ, BA, BB, BG, BR, (21) International Application Number: PCT/US99/23315 BY, CA, CH, CN, CU, CZ, DE, DK, EE, ES, FI, GB, GD. (22) International Filing Date: 6 October 1999 (06.10.99) GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, (30) Priority Data: SK, SL, TJ, TM, TR, TT, UA, UG, US, UZ, VN, YU, ZW, 09/169,289 9 October 1998 (09.10.98) US ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, (63) Related by Continuation (CON) or Continuation-in-Part ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI (CIP) to Earlier Application patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, US 09/169,289 (CIP) NE, SN, TD, TG). Filed on 9 October 1998 (09.10.98) Published (71) Applicant (for all designated States except US): INCYTE With international search report. PHARMACEUTICALS, INC. [US/US]; 3174 Porter Drive, Palo Alto, CA 94304 (US). (88) Date of publication of the international search report: 13 July 2000 (13.07.00) (72) Inventors; and (75) Inventors/Applicants (for US only): WALKER, Michael, G. [CA/US]; Unit 80, 1050 Borregas Avenue, Sunnyvale, CA 94089 (US). VOLKMUTH, Wayne [US/US]; 783 Roble Avenue, #1, Menlo Park, CA 94025 (US). KLINGLER, Tod, M. [US/US]; 28 Dover Court, San Carlos, CA 94070 (US). (74) Agents: BILLINGS, Lucy, J. et al.; Incyte Pharmaceuticals, Inc., 3174 Porter Drive, Palo Alto, CA 94304 (US).

(54) Title: MATRIX-REMODELING GENES

(57) Abstract

The invention provides novel matrix-remodeling genes and polypeptides encoded by those genes. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating or preventing diseases associated with matrix remodeling.

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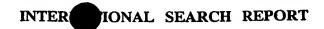
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Inter Pres Application No PCT/US 99/23315

A. CLASS	SIFICATION OF SUBJECT MATTER					
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Assording	to International Patent Classification (IDC)					
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Minimum d	ocumentation searched (classification system followed by classific	ation symbols)				
IPC 7	C12N C07K					
Documents	ation searched other than minimum documentation to the extent that	t such documents are included in the fields se	arched			
Electronic o	data base consulted during the international search (name of data t	pase and, where practical, search terms used				
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C. DOCUM	ENTS CONSIDERED TO BE RELEVANT					
Category °	Citation of document, with indication, where appropriate, of the r	elevant passages	Relevant to claim No.			
Α	MATRISIAN, L. M.: "METALLOPROT THEIR INHIBITORS IN MATRIX REMO	EINASES AND	1-12			
	TRENDS IN GENETICS,					
	vol. 6, no. 4, 1990, pages 121-	125,				
	XP000644304 * figure 2; page 123 *					
Α	YE, S. ET AL.: "MATRIX"		1-12			
	METALLOPROTEINASES:" CLINICAL SCIENCE.					
	vol. 94, February 1998 (1998-02)), pages				
	103-110, XP000857426					
	* whole disclosure *					
		-/				
		·				
X Furth	er documents are listed in the continuation of box C.	Patent family members are listed in	I annex.			
	egories of cited documents :	T later document published after the inten	national filing date			
"A" docume conside	nt defining the general state of the art which is not ered to be of particular relevance	cited to understand the principle or the	ne application but			
"E" earlier de	ocument but published on or after the international ate	"X" document of particular relevance: the claimed invention				
L documer which is	nt which may throw doubts on priority claim(s) or s cited to establish the publication date of another	involve an inventive step when the doc	pe considered to ument is taken alone			
citation or other special reason (as specified) Y document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the						
other means other means other means						
"P" document published prior to the international filing date but in the art. later than the priority date claimed "&" document member of the same patent family						
Date of the a	ctual completion of the international search	Date of mailing of the international sear	ch report			
27	January 2000	0 3, 05, 00				
Name and m	ailing address of the ISA	Authorized officer				
	European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel (*31-70) 340-2040 TV 31 651 cen el					
	Tel. (+31-70) 340-2040, Tx. 31 651 epp nl, Fax: (+31-70) 340-3016	HERMANN R.				



nter anal Application No PCT/US 99/23315

		PC1/05 99/23315
ategory °	ation) DOCUMENTS CONSIDERED TO BE RELEVANT Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Category	Onclusion of absument, with indication, where appropriate, or the relevant passages	nelevant to claim No.
Х	DATABASE EMBL/GENBANK [Online] ID HSZZ74011; AC AA368885, 18 April 1997 (1997-04-18) ADAMS, M.D. ET AL.: "EST80295 Placenta I homo sapiens cDNA" XP002128855 abstract	2-8
X	DATABASE EMBL/GENBANK [Online] ID HSW4574; AC W92457, 18 July 1996 (1996-07-18) HILLIER, L. ET AL.: "EST; Soares fetal heart NbHH19W homo sapiens cDNA clone" XP002128856 abstract	2-8
		-
	*	





ational application No. INTERNATIONAL SEARCH REPORT

PCT/US 99/23315

BoxI	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1. X	Claims Nos.: 10-12 because they relate to subject matter not required to be searched by this Authority, namely: Although claims 10-12 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. X	Claims Nos.: 2,7 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically: See FURTHER INFORMATION sheet PCT/ISA/210
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
	mational Searching Authority found multiple inventions in this international application, as follows:
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2.	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3.	As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.: 1-3,5-7,9,10,12 (all partially)
Remark o	The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.

International Application No. PCT/US 99 /23315

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 2,7

Claims 2 and 7 are -inter alia- drafted to compounds comprising as few as 18 nucleotides / 6 amino acids, which are neither defined by their exact structure, not by their exact location in the parent molecule. Moreover, claim 2 covers

- polynucleotides of completely undefined length that hybridize to said short sequence (or the parent), and

- sequences which are complementary to the almost undefined sequences mentioned above.

Said vague structural definitions are not sufficient for a reasonable search.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

1. Claims: 1-3,5-7,9,10,12 (all partially)

A polynucleotide having the SEQ ID NO 1; subject-matter of claims 2,3,5,6,9,12 relating to said polynucleotide

2. Claims: 1-12, all partially

Polypeptide with the SEQ ID NO. 21, encoding polynucleotides, and related subject-matter

3. Claims: 1-12, all partially

Polypeptide with the SEQ ID NO. 22, encoding polynucleotides, and related subject-matter

4. Claims: 1-12, all partially

Polypeptide with the SEQ ID NO. 23, encoding polynucleotides, and related subject-matter

5. Claims: 1-3,5-7,9,10,12 (all partially)

INVENTIONS 5-20: Polynucleotides having the SEQ ID NOs. 3-5,7-10,12-20; subject-matter of claims 2,3,5,6,9,12 relating to said polynucleotide.

6. Claims: 1,3 (partially)

INVENTION 21:

Further polynucleotides and encoded polypeptides of claim 1 (PLEASE NOTE: This subject has been added for completeness only. It is considered as being at least partially anticipated (infra); Further non-unity objections may arise upon paiment for subject 21)

